Update on gingival overgrowth by cyclosporine A in renal transplants

Domenico Ciavarella¹, Rosario Guiglia², Giuseppina Campisi², Michele Di Cosola³, Chiara Di Liberto², Antonio Sabatucci⁴, Nayra Escudero⁶, Antonio Bascones⁵, Lorenzo Lo Muzio¹

(1) Department of Surgical Sciences, University of Foggia, Foggia, Italy

- (2) Department of Dental Sciences, University of Palermo, Palermo, Italy
- (3) Department of Dentistry and Surgery, University of Bari, Bari, Italy
- (4) Institute of Dental Sciences, University of Ancona, Ancona, Italy
- (5) Department of Oral Medicine and Buccofacial Surgery, University Complutense, Madrid, Spain
- (6) Course of expert in clinical of periodontology. University Complutense, Madrid

Correspondence: Prof. Lorenzo Lo Muzio, Via Carelli 28 - 71100 Foggia – Italy. E-mail: lomuziol@tin.it

Received: 29-03-2006 Accepted: 20-05-2006 Ciavarella D, Guiglia R, Campisi G, Di Cosola M, Di Liberto C, Sabatucci A, Escudero N, Bascones A, Lo Muzio L. Update on gingival overgrowth by cyclosporine A in renal transplants. Med Oral Patol Oral Cir Bucal 2007;12:E19-25. © Medicina Oral S. L. C.I.F. B 96689336 - ISSN 1698-6946

ndexed in: -Index Medicus / MEDLINE / PubMed -EMBASE, Excerpta Medica -SCOPUS -Indice Médico Español -IBECS

ABSTRACT

Severe gingival overgrowth is one of the most frequent side effects in renal transplant patients associated with assumption of cyclosporine A. Several associations with age, sex, dosage, duration of therapy or interval since transplantation have been hypothesized. The introduction of alternative immunosuppressant drugs have been suggested to permit better long-term transplant outcomes and a decrease in incidence of gingival overgrowth. The aim of the present paper is to summarize current knowledge regarding aetiology, pathogenesis and management of gingival overgrowth induced by Cyclosporine A.

Key words: Gingival overgrowth, cyclosporine A, renal transplantation.

RESUMEN

El sobrecrecimiento gingival severo es uno de los efectos adversos más frecuentes en los pacientes con transplante renal asociado al suministro de ciclosporina A. Se han realizado hipótesis sobre diversas asociaciones con la edad, sexo, dosis, duración de la terapia o intervalo desde el transplante. Se ha propuesto la introducción de la alternativa de drogas inmunosupresoras para permitir mejores resultados a largo plazo del transplante y la disminución en la incidencia de sobrecrecimiento gingival. El objetivo del presente estudio es resumir el conocimiento actual, observando la etiología, patogénesis y dirección del sobrecrecimiento gingival inducido por la ciclosporina A.

Palabras clave: Sobrecrecimiento gingival, ciclosporina A, transplante renal.

INTRODUCTION

Organ transplantation procedures necessitate the use of immunosuppressant drugs. Immunosuppressant therapy is prescribed in treatment of autoimmune diseases and prevention of organ transplants rejection. The increased use of these medications has focused attention on the specific toxicities and side-effects associated (1, 2).

Since 1965, Merrill (3) performed in human beings the renal transplant and now it is considered a routine therapy for the treatment of irreversible renal failures. After renal transplantation, it is mandatory beginning immunosuppressant therapy in order to create a condition in which lymphocytes are not active, to avoid renal transplant rejection (4).

Cyclosporine A (CsA) has been the primary tool to prevent the rejection of organ transplants. The clinical use of CsA is often complicated by several well documented side effects including gingival overgrowth (GO) (5, 6) (Table 1).

The disfiguring GO may interfere with the normal oral functions and may also cause delayed and/or ectopic eruption of teeth, impaired speech, headache appearance and difficulty in maintaining optimal oral hygiene resulting in an increased susceptibility to infections, caries, and periodontal diseases (7). Furthermore, these consequences may have a psychological impact in children and may in turn influence compliance with medical therapy.

In the last years, new drugs have been used to avoid renal transplant rejection and to limit the well known side effects. Several studies indicate 'improvement' in GO when patients use new immunosuppressant drugs, such as Tacrolimus (FK506, Prograf® Fujisawa, Japan), Rapamycin RS 61443 and Mycophenolate mofetil (MMF) (CellCept®) (8), even if their collateral effects are not still extensively reported and, for this reason, CsA is still the mostly used drug in renal transplant therapy (9,10). However, there is evidence that use of Tacrolimus causes fewer oral side-effects than CsA (8,11,12).

INMUNOSUPPRESSANT THERAPY

After renal transplant, the therapeutic schemes of immunosuppressant drugs are several: at first time, it is used to give high dose of drugs followed to a support period in which reducing dosage therapeutic till the lowest dosage possible. The protocol applied has varied from mono-therapy, (6metilprednisolone or CsA) to a combination of two drugs (CsA and cortisone or alternatively CsA and dihydropyridine) up to the use of three drugs (CsA, cortisone and azathioprine) (9,10).

Among these drugs CsA is a non-myelotoxic immunosuppressant and its primary target is helper/inducer T lymphocytes; azathioprine is a non specific myelosuppressive commonly used together with prednisolone (triple medication), dihydropyridines are calcium-channel blocking agents used in the management of cardiovascular disorders, which have been shown to suppress cell-mediated immunity (13, 14). Dihydropyridines may have a synergistic immunosuppressant effect on CsA therapy (15) and on CsA induced suppression of T-cell proliferation (16) with the advantage of reducing the nephrotoxicity. Renal transplant patients are frequently treated with both CsA and dihydropyridine.

The increased use of immunosuppressive medication has focused attention on the specific toxicities and side-effects associated with these agents.

CYCLOSPORIN A

Cyclosporin (CsA) (Sandimmune® Novartis Pharmaceuticals Corporation, Hanover, Germany) was introduced in 1972 thanks to Borel of the Sandoz pharmaceutical company and registered in 1983. It is a lipophilic cyclic endecapeptide, isolated as an antifungal, from soil samples containing Cylindrocarpon lucidum BOOTH and Tolypocladium inflatum GAMS (fungi imperfecti) (17).

Nephrotoxicity	Hypertension
Hepatotoxicity	Biliary calculus disease
• Diabetes	Neurotoxicity
• Epilepsy	• Hirsutism
• Tremors	Altered bone metabolism
Lingual fungiform papillae hypertrophy	Gingival Overgrowth
Plasmocytoma	Kaposi's sarcoma
Squamous cell carcinoma of the lips	• Cephalalgy
• Sinusitis	Conjunctivitis
Hairy-leukoplakia	

Table 1.	Reported	side effects	of CsA	
Table 1.	reported	side chiects	01 03/1.	

The molecular tertiary structure results in the formation of hydrophilic immunosuppressant binding site and reduces immunosuppressant potential (18,19). CsA is variably absorbed in the gut and peak plasma concentration is reached after 3-4 hours. The drug is mostly bound to the following cells: 50% erythrocytes, 5% lymphocytes and 40% lipoproteins (20), with approximately 5% free in the plasma (7). CsA is metabolised in the liver microsomes (21), and excreted after 6 hours mainly via the bile, through the faeces (22).

In vitro and in vivo experiments indicated that CsA interferes selectively on T cell, particularly it inhibits T helper cells and it has get or no effect on T suppressor cells (23-25).

CsA inhibits selectively macrophage activation and IL-1 production, it prevents production of IL-1 receptors on T helper cells (26), inhibits IL-2 synthesis at low concentrations limiting clonal amplification of citotoxic T cell, and it inhibits their ability to respond to IL-2 (probably blocking cell surface receptors) (27).

CsA passively diffuses through cell membranes of many cells (including all peripheral blood lymphocytes and erytrocytes) and is concentrated in cytoplasm and nucleus (7).

The pharmacokinetic problems due to incomplete, unpredictable and inadequate absorption of original formulation of CsA have brought to introduction of new microemulsion formulation of CsA (Neoral), characterized to better absorption and a lower intra/inter-patient variability, permitting a improved long-term transplant outcomes (28, 29).

PATHOGENESIS

The pathogenesis of CsA-GO is probably multifactorial and still uncertain. Some studies have suggested associations between incidence-severity of GO and sex (30), age at transplantation (31, 32), duration of therapy and CsA dosage, (33-35) significant drug-related risk factors for the development of the clinical conditions (31, 32, 36-38). There is a wide intra and inter-individual variability in the susceptibility of the CsA to induce GO (CsA-GO) (range 30 to 50%) (39) and regularly it appears in more than 70% of adult transplant patients (31, 40).

It has been reported that CsA is able to alter the metabolism of human gingival fibroblasts (41) and the lamina propria extracellular components. Histologically, gingival hyperplasia is associated with an increase in the deposition of intercellular matrix, in the percentage of inflammatory cells (particularly macrophages, also known as Langerhans cells), and in the degree of tissue vascularization(42-46). This inflammatory response is increased by the presence of dental plaque, suggesting that the hyperplasia can represent a response to the bacterial toxins (47, 48).

Some studies have suggested that this type of GO is related to an individual drug or metabolite susceptibility (5) since gingival fibroblasts show an individual drug response (49). It is possible that CsA and its metabolites react with a phenotypically distinct subpopulation of gingival fibroblasts causing an increase in protein synthesis and rate of cell proliferation (50, 51). However, fibroblast heterogeneity has been demonstrated in relation to collagenase activity suggesting that CsA-GO involves an increase in the amount of collagen and extra cellular matrix formation (52) rather than an increase in the number of fibroblasts (53); this is could be a strange condition because IL-6 inhibits fibroblast proliferation (by the inhibiting of stimulatory cytokines) (54).

Morton et al. (55) found that gingival fibroblasts respond to CsA increasing IL-6 secretion, which itself enhances collagen and glycosaminoglycan synthesis. They also observed that CsA synergizes with IL-1B to further up-regulated IL-6 secretion asserted that one of the pathogenic mechanisms underlying drug-induced GO might be enhanced secretion of IL-6 by gingival fibroblasts (55). It has been reported that gingival fibroblasts produce considerable quantity of IL-6, especially after stimulation with bacteria or other cytokines (56).

In renal transplantation patients (57) increased levels of IL-6 have been reported, (58,59), but the cellular origin of IL-6 in CsA-GO is unknown. Although immune cells such as lymphocytes and monocytes could be the source of this cytokine, it is evident that the substantial rise in IL-6 activity in renal transplant or gingival tissues of CsA-treated patients cannot be attributed to inflammatory cell infiltrates (60, 61).

In relation to phagocytosis, fibroblasts from CsA-GO showed reduced phagocyte activity in inflamed tissue, as a result of lower proportions of phagocytes (35, 62). An increased rate of synthesis coupled with a decreased rate of phagocytosis would result in an evident increased amount of essential substance and it could explain the increase of connective tissue volume in CsA-GO.

Hence, it is possible to hypothesize that those individuals treated by CsA and genetically susceptible to GO, have got an appropriate mixture of fibroblast and lymphocyte subpopulations, as being able to interact with CsA in inflamed gingival tissue, up to the well known GO clinical manifestations (63).

CSA THERAPY IN AUTOINMUNE DISORDERS AND ORAL SIDE EFFECTS

CsA is increasingly used, not only in transplantation practice but, also, in autoimmune disorders (i.e. psoriasis, systemic lupus erythematous, rheumatoid arthritis and multiple sclerosis) causing equally various side effects as in the oral cavity. In this district, the rare lingual fungiform papillae hypertrophy (LFPH), GO (64, 65), opportunistic infections, hairy leukoplakia (66, 67) and squamous cells carcinoma of the lip (67) have been repeatedly reported. Many studies have examined interaction of CsA with gingival fibroblasts, reporting a correlation between CsA effects on fibroblasts and extra-cellular components of connective lamina propria and GO; in fact, gingival fibroblasts show an individual response to the drug (68). Clinically gingival hyperplasia begins at the interdental papillae, more commonly in anterior than in posterior region, and frequently on labial than on lingual surfaces (30, 69, 70).

Hyperplasia is often limited on adherent gingiva, but it may to extend coronally and interfere with occlusion, mastication and language, without necessarily altering the underlying periodontium. Hyperplasia origin is apparently due to many factors: age, dosage, time of CsA assumption and possible additional other drugs. CsA-GO has not been described in edentulous patients or in edentulous space (40,71), even if few authors have reported CsA-GO in edentulous patients (71-75); in these latter cases an association of the hyperplasia emerges with denture trauma or candidal infection (71).

RISK FACTORS ASSOCIATED WITH SEVERITY OF GINGIVAL OVERGROWTH

Serum and salivary concentrations, dosage of the drug, time since transplantation and assumption of drug, age, concomitant medication and oral hygiene are some the main risk factors influencing the severity of GO in renal transplantation patients.

SERUM CONCENTRATION AND DOSAGE

This point has been discussed in controversial manner: some authors have found a correlation between the dosage of CsA and development severity of GO (34, 76), while others have reported contrasting findings (7, 42, 78, 79). There is a general agreement that an initial threshold serum concentration is required to initiate GO and it has been suggested that the higher is the serum concentration the quicker GO develops (76).

SALIVARY CONCENTRATION

Salivary concentrations of CsA are higher in patients taking the liquid form of the drug compared to the capsule form, but salivary concentrations are poorly correlated with blood levels (80).

TIME OF ADMINISTRATION

The relationship between GO and time of administration is another controversial issue since GO has been reported both in patients taking CsA within three months (34) or more (5). But, a recent report in children indicated that only subjects taking CsA for more than three months showed in all cases GO (81).

DURATION

Duration of CsA therapy has been reported to increase the chance to have GO. Recently, a significant inverse correlation between periodontal status and duration of therapy has been reported, thus the negative effects of the drug could spontaneously decrease over time (82).

AGE AND SEX

Prevalence and severity of GO has been reported to be significantly higher in children in comparison to the adults undergoing organ transplants. (79, 83). The immature fibroblasts, probably, are more sensitive to the effect of CsA (84, 85), so the increase GO in adolescents can be due to an interaction between circulating androgens, estrogens and gingival fibroblasts. Studies seem to suggest that children, especially adolescents and females may be more susceptible to this undesirable effect than adults (42, 86) The high-level of sexual hormones found in patients of this age produce an active metabolite (5 alpha-dihydrotestosterone) that acts on a subpopulation of gingival fibroblasts as being to the increase collagen synthesis and/or to decrease the collagenase production (87). In addition to the potential effect of hormones in adolescence, there is evidence that GO is influenced by orthodontic appliances which may irritate the soft tissues and act as a plaque trap thereby increasing the GO (88).

COMBINATIONS OF MEDICATIONS ASSOCIA-TED WITH GINGIVAL OVERGROWTH

The most common combination of drugs that cause GO is CsA and nifedipine. Nifedipine is used habitually to treat hypertension which may be primary or secondary to CsA nephrotoxicity (36, 89). A significant increase in the incidence of GO has been described in renal transplant patients taking nifedipine and CsA compared with those taking CsA alone (51 per cent vs 8 per cent) (90).

GENETIC PREDISPOSITION

The role of individual susceptibility to GO has recently attracted much attention. Several studies (32, 37, 38) have described slight disease association between HLA alleles (A19, A24 and DR-2), and a possible protective effect of HLA-B37 and DR-1. The selectivity of the overgrowth to particular regions of the mouth and only in some patients, suggested a genetic predisposition that may interact with the local environment resulting in GO (39).

ORAL HYGIENE

It is well known that dental plaque is composed of an aggregation of micro organisms deposited on the tooth surfaces and it is able to begin inflammatory changes in the gingival tissues. Mediators of the inflammatory response cause the changes in gingival tissues (91). These inflammatory changes may increase the interaction between CsA and fibroblasts leading to changes in connective tissue turnover. The improvement of general standard of oral hygiene can reduce GO resolving the inflammatory component plaque induced. In a longitudinal study, the effect on GO-CsA of a given oral hygiene program was examined in adult transplant patients (92): both the oral hygiene and the control group registered a significant gingival change, however, the degree of GO was less marked in the group with oral hygiene program.

Although the role of plaque has not been clearly defined consistently with other several authors (93-96), the hyperplastic tissue tends to aid plaque accumulation and to inhibit plaque removal, increases the gingival inflammation.

MANAGEMENT OF GINGIVAL OVER-GROWTH

Different therapeutic approaches for GO management have been proposed. The use of specific oral hygiene programs, surgical intervention and/or alternative pharmacological therapy have been reported.

Severe oral hygiene measures may reduce the degree of GO, but they do not inhibit its development.

Reduction in the dose of CsA has been shown to be beneficial versus GO but, frequently, the nature of organ transplants does not permit an alternative therapy or dose reduction (97).

When possible, alternatively to CsA, some patients can use other immunosuppressant drugs such as tacrolimus (FK506), azathioprine and rapamycin that may offer some hope not reporting GO associated. There is evidence to support the use of Tacrolimus over CsA since it has shown to cause fewer oral side-effects than CsA (11, 12), even if tacrolimus may stimulate formation of cyst in anterior site of mouth and increase infections by Human Papilloma Virus, xerostomia, and oral ulcers (67). Tacrolimus/FK 506, a macrolide molecule, acts primarily on CD4+ T helper lymphocytes by inhibiting the production of lymphokines. which are required for cell growth and differentiation, principally of IL-2, at the transcriptional level (98). It has also been suggested that the elimination of CsA and its replacement of Tacrolimus removes the up regulation of CsA-induced essential polypeptide growth factors, which are important mediators in development of GO (99).

CONCLUSION

CsA is an immunosuppressant used widely for its rejectionpreventing power in human organ of transplantation (7, 100). One of the side effects associated with CsA treatment is GO; CsA-GO is generally associated with acute or chronic inflammation due to plaque accumulation (101, 102), reported dosage of CsA (103), presence of local gingival inflammation and gingivitis as independent predictors of the extent/severity of GO. An individual genetic susceptibility can be more or less responsible of the clinical manifestation of GO.

The present paper is drawn to underline the etiological relationships and clarify some important aspects on the risk factors responsible of CsA-GO in renal transplantation patients. In the future, dosage reduction and/or replacement of CsA with alternative efficacious pharmacological therapies to the CsA will undoubtedly taken into account in order to improve the oral health in these patients.

REFERENCES

1. Vella JP, Sayegh MH. Developments in the clinical science of transplantation during the 20th century. Pediatr Transplant 1998;2: 257-62.

2. Denton MD, Magee CC. Immunosuppressive strategies in transplantation. Lancet 1999;353:1083-91.

3. Merrill J P, Murray JE. Successful homotransplantation of the human kidney between identical twins. J Am Med Assoc 1956;160: 277-82.

4. Land W. Kidney transplantation-state of the art. Transplant Proc 1989;21:1425-9.

5. Wysocki GP, Gretzinger HA. Fibrous hyperplasia of the gingiva: a side effect of cyclosporin A therapy. Oral Surg Oral Med Oral Pathol 1983;55:274-8.

6. Paul LC. Overview of side effects of immunosuppressive therapy. Transplant Proc 2001;33:2089-91.

7. Hassell T M, Hefti AF. Drug-induced gingival overgrowth: old problem, new problem. Crit Rev Oral Biol Med 1991;2:103-37.

8. Budde K, Fritsche L. Clinical pharmacokinetics of tacrolimus in rescue therapy after renal transplantation. Int J Clin Pharmacol Ther 1996;34:493-7.

9. Ponticelli C, Tarantino A. From cyclosporine to the future. Transplant Proc 2004;36:557S-560S.

10. Verma A, Dhawan A. Re: Trial of metronidazole vs. azithromycin for treatment of cyclosporine-induced gingival overgrowth. Pediatr Transplant 2005;9:132.

11. Cox KL, Freese DK. Tacrolimus (FK506): the pros and cons of its use as an immunosuppressant in pediatric liver transplantation. Clin Invest Med 1996;19:389-92.

12. Hernández G, Arriba L. Reduction of severe gingival overgrowth in a kidney transplant patient by replacing cyclosporin A with tacrolimus.J Periodontol 2000;71:1630-6.

13. Solovera JJ, Alvarez-Mon M. Inhibition of human natural killer (NK) activity by calcium channel modulators and a calmodulin antagonist. J Immunol 1987;139:876-80.

14. Morgano A, Pierri I. Decreased lymphocyte blastogenesis, IL2 production and NK activity following nifedipine administration to healthy humans. Eur J Clin Pharmacol 1990;39:545-50.

15. Padberg WM, Bodewig C. Synergistic immunosuppressive effect of low-dose cyclosporine A and the calcium antagonist nifedipine, mediated by the generation of suppressor cells. Transplant Proc 1990;22:2337.

16. Marx M, Weber M. Additive effects of calcium antagonists on cyclosporin A-induced inhibition of T-cell proliferation. Nephrol Dial Transplant 1990;5:1038-44.

17. Dreyfuss, M, Haerri E. Cyclosporine A and C. New metabolites from Trichoderma polysporum. Eur J Appl Microbiol 1976;3:125-33.

18. Wenger RM. Synthesis of ciclosporin and analogues: structural and conformational requirements for immunosuppressive activity. Prog Allergy 1986;38:46-64.

19. Wenger RM. Cyclosporine: conformation and analogues as tools for studying its mechanism of action. Transplant Proc 1998;20:313-8.

20. Rodl S, Khoshsorur G. Binding of cyclosporine A to human serum lipoproteins. Transplant Proc 1990;22:287-8.

21. Combalbert J, Fabre I. Metabolism of cyclosporin A. IV. Purification and identification of the rifampicin-inducible human liver cytochrome P-450 (cyclosporin A oxidase) as a product of P450IIIA gene subfamily. Drug Metab Dispos 1989;17:197-207.

22. Venkataramanan R, Starzl TE. Biliary excretion of cyclosporin in liver transplant patients. Transpl Proc 1985;17:286-9.

23. Gao EK, Lo D. Abnormal differentiation of thymocytes in mice treated with cyclosporin A. Nature 1988;336:176-9.

24. Jenkins MK, Schwartz RH. (1988). Effects of cyclosporine A on T cell development and clonal deletion Science 1988;241:1655-8.

25. Thomson AW, Webster LM. The influence of cyclosporin A on cellmediated immunity. Clin Exp Immunol 1988;71:369-76.

26. Thomson A, Moon D. Cyclosporine and lymphokines affecting macrophage behaviour. Trans Proc 1983;15:2390-3.

27. Hess AD, Colombani PM. Mechanism of action of cyclosporine: a unifying hypothesis. Adv Exp Med Biol 1987;213:309-30.

28. Mueller EA, Kovarik JM. Improved dose linearity of cyclosporine pharmacokinetics from a microemulsion formulation. Pharm Res 1994;11:301-4.

29. Grant D, Kneteman N. Peak cyclosporine levels (Cmax) correlate with freedom from liver graft rejection: results of a prospective, randomized comparison of neoral and sandimmune for liver transplantation (NOF-8). Transplantation 1999;67:1133-7.

30. Tyldesley W, Rotter E. Gingival hyperplasia induced by cyclosporin A. Br Dent J 1984;157:305-9.

31. Daley TD, Wysocki GP. Clinical and pharmacologic correlations in cyclosporine-induced gingival hyperplasia. Oral Surg Oral Med Oral Pathol 1986;62:417-21.

32. Margiotta V, Pizzo I. Cyclosporin- and nifedipine-induced gingival overgrowth in renal transplant patients: correlations with periodontal and pharmacological parameters, and HLA-antigens. J Oral Pathol Med 1996;25:128-34.

33. Hefti AF, Eshenaur AE. Gingival overgrowth in cyclosporine A treated multiple sclerosis patients. J Periodontol 1994;65:744-9.

34. Seymour R A, Smith DG. The comparative effects of azathioprine and cyclosporin on some gingival health parameters of renal transplant patients. A longitudinal study. J Clin Periodontol 1987;14:610-3.

35. McGaw WT, Porter H. Cyclosporine-induced gingival overgrowth: an ultrastructural stereologic study. Oral Surg Oral Med Oral Pathol 1998;65:186-90.

36. Thomason JM, Seymour RA. The prevalence and severity of cyclosporin and nifedipine-induced gingival overgrowth. J Clin Periodontol 1993;20:37-40.

37. Pernu HE, Pernu LM. Gingival overgrowth among renal transplant recipients related to immunosuppressive medication and possible local background factors. J Periodontol 1992;63:548-53.

38. Thomason JM, Seymour RA. Determinants of gingival overgrowth severity in organ transplant patients. An examination of the role of HLA phenotype. J Clin 1996;23:628-34.

39. Seymour RA, Thomason JM. The pathogenesis of drug-induced gingival overgrowth. J Clin Periodontol 1996;23:165-75.

40. Friskopp J, Klintmalm G. Gingival enlargement. A comparison between cyclosporine and azathioprine treated renal allograft recipients. Swed Dent J 1986;10:85-92.

41. Tipton DA, Stricklin GP. Fibroblast heterogeneity in collagenolytic response to cyclosporine. J Cell Biochem 1991;46:152-65.

42. Seymour RA, Jacobs DJ. Cyclosporin and the gingival tissues. J Clin Periodontol 1992;19:1-11.

43. Kinane DF, Drummond JR. Langerhans cells in human chronic gingivitis and phenytoin-induced gingival hyperplasia. Arch Oral Biol 1990;35:561-4.

44. Brown RS, Di Stanislao PT. The administration of folic acid to institutionalized epileptic adults with phenytoin-induced gingival hyperplasia. A double-blind, randomized, placebo-controlled, parallel study. Oral Surg Oral Med Oral Pathol 1991;71:565-8.

45. Brown RS, Beaver WT. On the mechanism of drug-induced gingival hyperplasia. J Oral Pathol Med 1991;20:201-9.

46. Romanos GE, Schroter-Kermani C. Extracellular matrix analysis of nifedipine-induced gingival overgrowth: immunohistochemical distribution of different collagen types as well as the glycoprotein fibronectin.J Periodontal Res 1993;28:10-6.

47. Penarrocha-Diago M, Bagan-Sebastian JV. Diphenylhydantoin-induced gingival overgrowth in man: a clinico-pathological study. J Periodontol 1990;61:571-4.

48. Hancock RH, Swan RH. Nifedipine-induced gingival overgrowth. Report of a case treated by controlling plaque. J Clin Periodontol 1992; 19:12-4.

49. Hassell TM, Romberg E. Lymphocyte-mediated effects of cyclosporine on human fibroblasts. Transplant Proc 1988;20:993-1002.

50. Hassell TM, Buchanan J. Fluorescence activated vital cell sorting of human fibroblast subpopulations that bind Cyclosporin A. Journal of Dental Research 1988;67:273.

51. Jacobs D, Hassell TM. The effect of cyclosporin metabolite OL-17 on gingival fibroblast sub populations. Journal of Dental Research 1990; 69:221.

52. Williamson MS, Miller EK. Cyclosporine A upregulates interleukin-6 gene expression in human gingiva: possible mechanism for gingival overgrowth. J Periodontol 1994;65:895-903.

53. Butler RT, Kalkwarf KL. Drug-induced gingival hyperplasia: phenytoin, cyclosporine, and nifedipine. J Am Dent Assoc 1987;114: 56-60.

54. Myrillas TT, Linden GJ. Cyclosporin A regulates interleukin-1beta and interleukin-6 expression in gingiva: implications for gingival overgrowth. J Periodontol 1999;70:294-300.

55. Morton RS, Dongari-Bagtzoglou AI. Regulation of gingival fibroblast interleukin-6 secretion by cyclosporine A. J Periodontol 1999;70: 1464-71.

56. Dongari-Bagtzoglou AI, Ebersole JL. Production of inflammatory mediators and cytokines by human gingival fibroblasts following bacterial challenge. J Periodontal Res 1996;31:90-8.

57. Joseph JV, Guy SP. Th1 and Th2 cytokine gene expression in human renal allografts. Transplant Proc 1995;27:915-6.

58. Van Oers MH, Van der Heyden AA. Interleukin 6 (IL-6) in serum and urine of renal transplant recipients. Clin Exp Immunol 1988; 71:314-9.

59. Raasveld MH, Bloemena E. Interleukin-6 and neopterin in renal transplant recipients: a longitudinal study. Transpl Int 1993;6:89-94.

60. Ford HR, Hoffman RA. Evidence that production of interleukin 6 within the rejecting allograft coincides with cytotoxic T lymphocyte development. Transplantation 1991;51:656-61.

61. Iacopino AM, Doxey D. Phenytoin and cyclosporine A specifically regulate macrophage phenotype and expression of platelet-derived growth factor and interleukin-1 in vitro and in vivo: possible molecular mechanism of drug-induced gingival hyperplasia. J Periodontol 1997;68:73-83.

62. McCulloch CA, Knowles GC. Deficiencies in collagen phagocytosis by human fibroblasts in vitro: a mechanism for fibrosis? J Cell Physiol 1993;155:461-71.

63. Pernu HE, Knuuttila ML. Drug-induced gingival overgrowth and class II major histocompatibility antigens. Transplantation 1994; 57:1811-3.

64. Starzl TE, Weil R, 3rd. The use of cyclosporin A and prednisone in cadaver kidney transplantation. Surg Gynecol Obstet 1980;151:17-26.

65. Rateitschak-Pluss EM, Hefti A. Initial observation that cyclosporin-A induces gingival enlargement in man. J Clin Periodontol 1983; 10:237-46.

66. King GN, Healy CM. Prevalence and risk factors associated with leukoplakia, hairy leukoplakia, erythematous candidiasis, and gingival hyperplasia in renal transplant recipients. Oral Surg Oral Med Oral Pathol 1994;78:718-26.

67. Seymour RA, Thomason JM. Oral lesions in organ transplant patients. J Oral Pathol Med 1997;26:297-304.

68. Hassell TM, Gilbert GH. Phenytoin sensitivity of fibroblasts as the basis for susceptibility to gingival enlargement. Am J Pathol 1983;112: 218-23.

69. Daley T D, Wysocki GP. Cyclosporin Therapy. Its significance to the periodontist. J Periodontol 1984;55:708-12.

70. Gonzalez-Jaranay Ruiz M, Mesa Aguado F. Cyclosporin induced gingival hyperplasia. Rev Eur Odontoestomatol 1991;3:265-70.

71. Thomason JM, Seymour RA. Severe mucosal hyperplasia of the edentulous maxilla associated with immunosuppressant therapy: a clinical report. J Prosthet Dent 1994;72:1-3.

72. Axell T. A prevalence study of oral mucosal lesions in an adult Swedish population. Odontol Revy 1976;27:1-103.

73. Dreyer WP, Thomas CJ. Diphenylhydantoinate-induced hyperplasia of the masticatory mucosa in an edentulous epileptic patient. Oral Surg Oral Med Oral Pathol 1978 45:701-6.

74. Bredfeldt G. W. Phenytoin-induced hyperplasia found in edentulous patients. J Am Dent Assoc 1992;123:61-4.

75. McCord JF, Sloan P. Phenytoin hyperplasia occurring under complete dentures: a clinical report. J Prosthet Dent 192;68:569-72.

76. Somacarrera ML, Hernandez G. Factors related to the incidence and severity of cyclosporin-induced gingival overgrowth in transplant patients. A longitudinal study. J Periodontol 1994;65:671-5.

77. Seymour RA, Heasman PA. Drugs and the periodontium. J Clin Periodontol 1988;15:1-16.

78. Lowry LY, Welbury RR. Gingival overgrowth in paediatric cardiac transplant patients: a study of 19 patients aged between 2 and 16 years. Int J Paediatr Dent 1995;54:217-22.

79. Hosey MT, Gordon G. Oral findings in children with liver transplants. International Journal of Paediatric Dentistry 1955;5:29-34.

80. Modeer T, Wondimu B. Levels of cyclosporin-A (CsA) in saliva in children after oral administration of the drug in mixture or in capsule form. Scand J Dent Res 1992;100:366-70.

81. Karpinia KA, Matt M. Factors affecting cyclosporine-induced gingival overgrowth in pediatric renal transplant recipients. Pediatr Dent 1996;18:450-5.

82. Montebugnoli L, Servidio D. The role of time in reducing gingival overgrowth in heart-transplanted patients following cyclosporin therapy. J Clin Periodontol 2000;27:611-4.

83. Kilpatrick NM, Weintraub RG. Gingival overgrowth in pediatric heart and heart-lung transplant recipients. J Heart Lung Transplant 1997; 16:1231-7.

84. Ross PJ, Nazif MM. Effects of Cyclosporin A on gingival status following liver transplantation. ASDC J Dent Child 1989;56:56-9.

85. Morisaki I, Kitamura K. Age dependency of cyclosporin A-induced gingival overgrowth in rats. Pediatr Dent 1993;15:414-7.

86. Hassell TM. Evidence for production of an inactive collagenase

by fibroblasts from phenytoin-enlarged human gingivae. J Oral Pathol 1982;11:310-7.

87. Sooriyamoorthy M, Gower DB. Androgen metabolism in gingival hyperplasia induced by nifedipine and cyclosporin. J Periodontal Res 1990;25:25-30.

88. Daley TD, Wysocki GP. Orthodontic therapy in the patient treated with cyclosporine. Am J Orthod Dentofacial Orthop 1991;100:537-41.

89. O'Valle F, Mesa F. Gingival overgrowth induced by nifedipine and cyclosporin A. Clinical and morphometric study with image analysis. J Clin Periodontol 1995;22:591-7.

90. Slavin J, Taylor J. Cyclosporin, nifedipine, and gingival hyperplasia. Lancet 1987;2:739.

91. Varga E, Lennon MA. Pre-transplant gingival hyperplasia predicts severe cyclosporin-induced gingival overgrowth in renal transplant patients. J Clin Periodontol 1998;25:225-30.

92. Seymour RA, Smith DG. The effect of a plaque control programme on the incidence and severity of cyclosporin-induced gingival changes. J Clin Periodontol 1991;18:107-10.

93. Steinberg SC. and Steinberg AD. Phenytoin-induced gingival overgrowth control in severely retarded children. J Periodontol 1982;53: 429-33.

94. Addy V, McElnay JC. Risk factors in phenytoin-induced gingival hyperplasia. J Periodontol 1983;54:373-7.

95. Pihlstrom BL, Carlson JF. Prevention of phenytoin associated gingival enlargement--a 15-month longitudinal study. J Periodontol 1980; 51:311-7.

96. Modeer T, Dahllof G. Development of phenytoin-induced gingival overgrowth in non-institutionalized epileptic children subjected to different plaque control programs. Acta Odontol Scand 1987;45:81-5.

97. Daly C. Resolution of cyclosporin A (CsA)-induced gingival enlargment following reduction in CsA dosage. J Clin Periodontol 1992;19:143-5.

98. Letko E, Bhol K. Tacrolimus (FK 506). Ann Allergy Asthma Immunol 1999;83: 179-89; quiz 189-90.

99. Boltchi FE, Rees TD.Cyclosporine A-induced gingival overgrowth: a comprehensive review. Quintessence Int1999;30:775-83.

100. Grune S. Proceedings of the Second International Congress on Cyclosporine. Washington DC. Philadelphia. 1988

101. Fu E, Nieh S. The effect of plaque retention on cyclosporine-induced gingival overgrowth in rats. J Periodontol 1997;68:92-8.

102. Page RC. The role of inflammatory mediators in the pathogenesis of periodontal disease. J Periodontal Res 1991;26:230-42.

103. Thomas DW, Newcombe RG. Risk factors in the development of cyclosporine-induced gingival overgrowth. Transplantation 2000; 69: 522-6.